

LSBio™ Mouse/Human Dysferlin Cell-Based ELISA Kit

Catalog No. LS-F1949

User Manual

**Please Read the Manual Carefully
Before Starting your Experiment**



LSBio

For research use only. Not approved for use in humans or for clinical diagnosis.

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INTRODUCTION

Colorimetric Cell-Based ELISAs

The Colorimetric Cell-Based ELISA Kit allows for the detection of various target proteins and the effects that certain stimulation conditions have on target protein expression in different cell lines. Qualitative determination of target protein concentration is achieved by an indirect ELISA format. In essence, the target protein is captured by target-specific primary (1°) antibodies while the HRP-conjugated secondary (2°) antibodies bind the Fc region of the 1° antibody. Through this binding, the HRP enzyme conjugated to the 2° antibody can catalyze a colorimetric reaction upon substrate addition. Due to the qualitative nature of the Cell-Based ELISA, multiple normalization methods are described: 1) a monoclonal antibody specific for human GAPDH is included to serve as an internal positive control in normalizing the target absorbance values. 2) Following the colorimetric measurement of HRP activity via substrate addition, the Crystal Violet whole-cell staining method is used to determine cell density. After staining, the results can be analyzed by normalizing the absorbance values to cell amounts, by which the plating difference can be adjusted. 3) If a phosphorylated target is being detected, an antibody against the non-phosphorylated counterpart will be provided for normalization purposes. The absorbance values obtained for the non-phosphorylated target can be used to normalize the absorbance values for the phosphorylated target.

Dysferlin Colorimetric Cell-Based ELISA

The Dysferlin Cell-Based ELISA Kit is a convenient, lysate-free, high throughput and sensitive assay kit that can monitor Dysferlin protein expression profile in cells. The kit can be used for measuring the relative amounts of Dysferlin in cultured cells as well as screening for the effects that various treatments, inhibitors (ie. siRNA or chemicals), or activators have on Dysferlin.

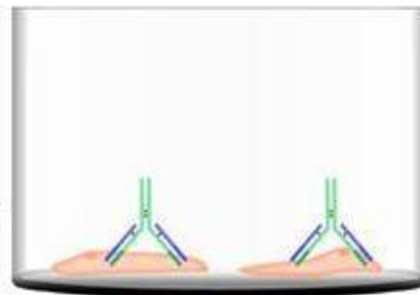
ASSAY FORMAT


**α-Target
Primary Antibody**



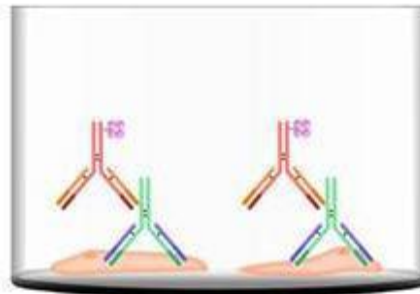
Cells are seeded onto the bottom of each well. The cells are quenched, fixed and the well is blocked.


**α-Primary IgG
Secondary Antibody**



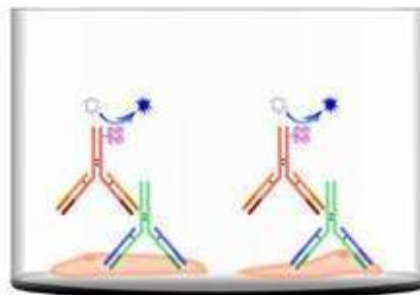
Primary antibodies specific for the target antigen are added and allowed to bind to their respective epitopes.



HRP Conjugate



HRP-conjugated secondary antibodies specific for the primary antibody are added and allowed to bind to their respective epitopes.


Unreacted TMB




**Blue TMB
Diimine Product**

TMB substrate is converted to the blue TMB diimine via the HRP enzyme. Upon addition of acid, the reaction terminates and the wells can be read at 450 nm.

ASSAY RESTRICTIONS

- This ELISA kit is intended for research purposes only, NOT diagnostic or clinical procedures of any kind.
- Materials included in this kit should NOT be used past the expiration date on the kit label.
- Reagents or substrates included in this kit should NOT be mixed or substituted with reagents or substrates from any other kits.
- Variations in pipetting technique, washing technique, operator laboratory technique, kit age, incubation time or temperature may cause differences in binding affinity of the materials provided.
- The assay is designed to eliminate interference and background by other cellular macromolecules or factors present within any biological samples. However, the possibility of background noise cannot be fully excluded until all factors have been tested using the assay kit.

SAMPLE COPY

ANTIBODY SPECIFICITY

Anti-Dysferlin Antibody

The Anti-Dysferlin Antibody is a rabbit polyclonal antibody. It was tested on Western Blots for specificity. The data in Figure 2 shows that a single protein band was detected. This protein band can be blocked by the synthesized immunogen peptide.

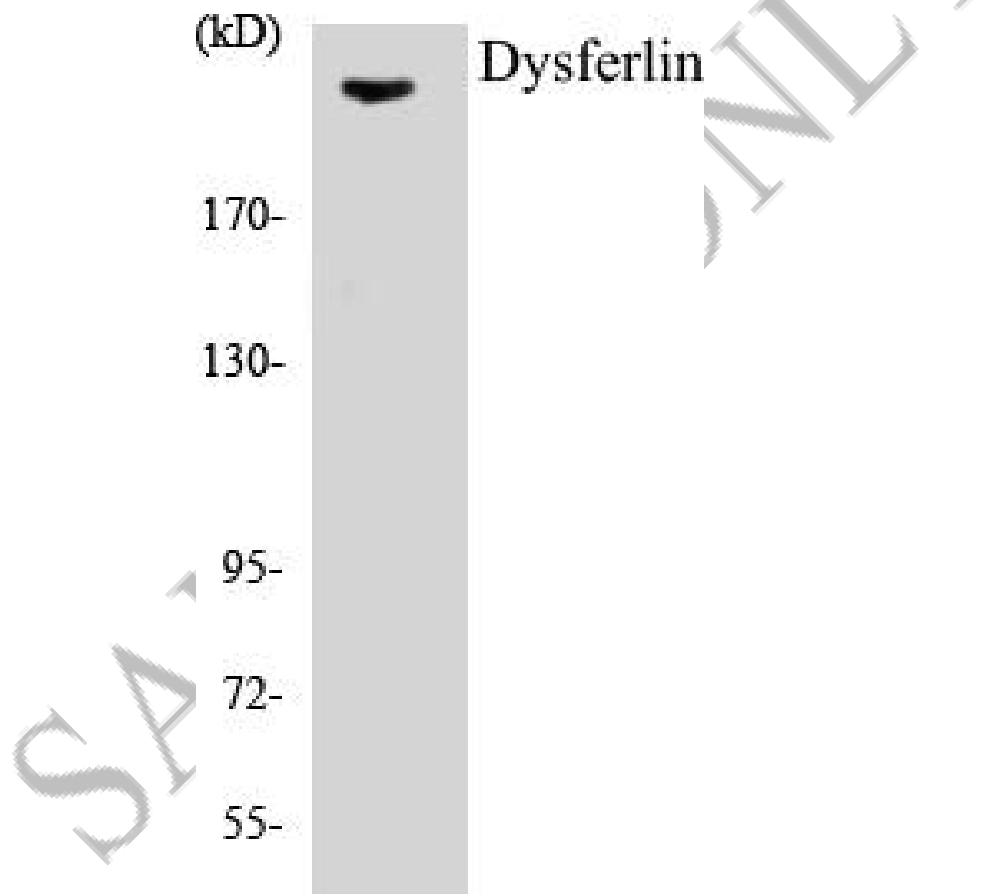


Figure 2. Western blot analysis of extracts from K562 cells, using Dysferlin Antibody.

Anti-GAPDH Antibody

The Anti-GAPDH Antibody is a mouse monoclonal antibody. It was tested on Western Blots with the tissue lysates from human, mouse, and rat for specificity. The data in Figure 3 shows that a single protein band was detected from all three lysates.

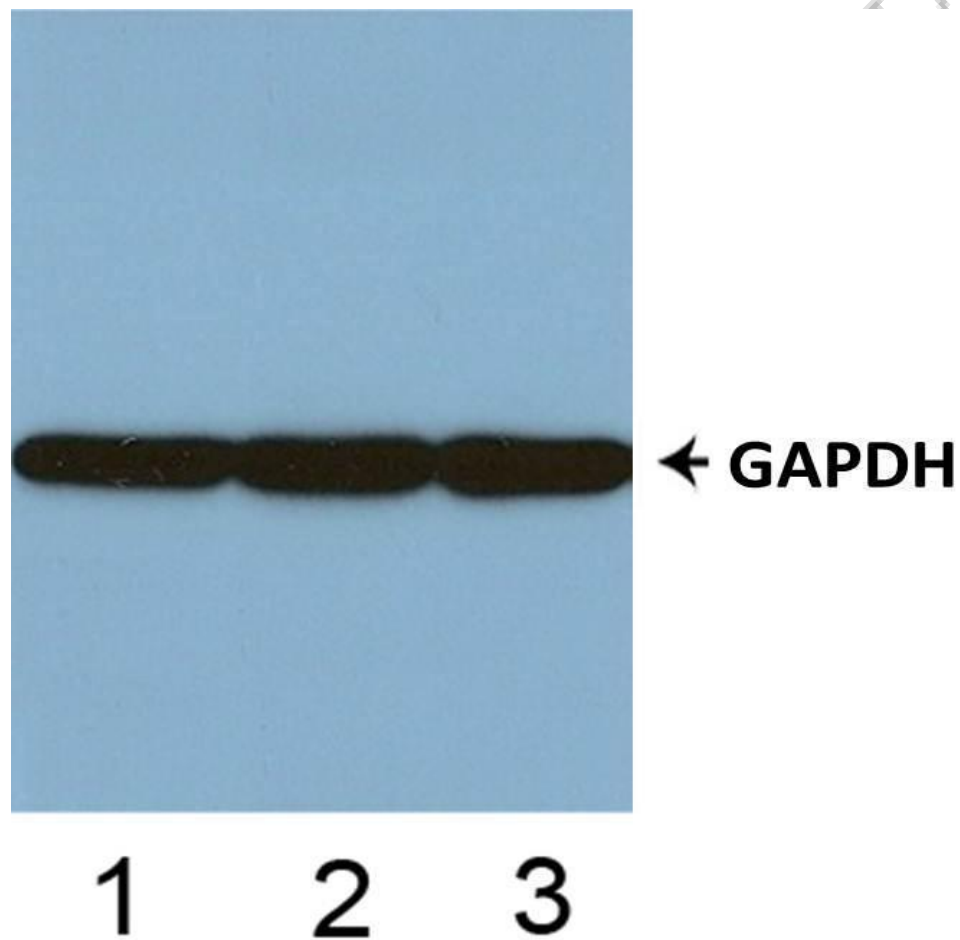


Figure 3. Western blot analysis of tissue lysates from human (1), mouse (2) and rat (3).

MATERIALS INCLUDED

Reagent	Quantity	Storage/Stability after first use
96-Well Cell Culture Clear-Bottom Microplate	1 Plate	-
10x TBS	24 ml (10x)	1 month at 4°C
Quenching Buffer	24 ml (1x)	
Blocking Buffer	50 ml (1x)	
15x Wash Buffer	50 ml (15x)	
100x Anti- Dysferlin Antibody	60 µl (100x)	
100x Anti-GAPDH Antibody	60 µl (100x)	
HRP-Conjugated Anti-Rabbit IgG Antibody	6 ml (1x)	
HRP-Conjugated Anti-Mouse IgG Antibody	6 ml (1x)	
Primary Antibody Diluent	12 ml (1x)	
Ready to Use Substrate	12 ml (1x)	
Stop Solution	12 ml (1x)	
Crystal Violet Solution	6 ml (1x)	
SDS Solution	24 ml (1x)	
Adhesive Plate Seals	2 Seals	

STORAGE AND STABILITY

Upon receipt, the kit should be stored at 4°C. The un-opened kit will be stable for up to 6 months from the date of shipment if stored at 4°C.

REAGENT PREPARATION

Ensure that all supplied solutions are at ambient temperature before use. It is recommended to conduct assays for all controls and samples in duplicate. Sufficient reagents are provided to assay 96 wells; therefore, on the day of the experiment, prepare only the required amount. During incubation steps, use an orbital shaker set at 200 rpm to ensure proper equilibration of solutions in the cell culture plate wells. **All other provided solutions are ready to use.**

1. Dilute the 10x TBS to 1x TBS by combining 9 volumes of ddH₂O and 1 volume of 10x TBS.
2. Dilute the 15x Wash Buffer to 1x Wash Buffer using 14 volumes of ddH₂O and 1 volume of 15x Wash Buffer.
3. Prepare the Fixing solution to 4% paraformaldehyde (for adherent cell lines) or 8% paraformaldehyde (for suspension or loosely attaching cell lines). Dilute 37% paraformaldehyde with 1x PBS to desired paraformaldehyde percentage.
4. For suspension or loosely attached cells: Prepare Poly-L-Lysine to working concentration as stated in manual. (Not included, recommended Sigma Cat# P4832).

If the cell line is known to digest Poly-L-Lysine, Poly-D-Lysine may be an alternative for this step. Follow the manufacturer's instructions if using a different Poly-L-Lysine or Poly-D-Lysine.

5. Prepare a 1x dilution of both 100x primary antibodies anti-Dysferlin and Anti-GAPDH by using the Primary Antibody Diluent at a ratio of 1:100.

ADDITIONAL MATERIALS REQUIRED

The following materials and equipment are NOT provided in this kit but are necessary to successfully conduct the experiment:

- Microplate reader able to measure absorbance at 450 nm and/or 595 nm for Crystal Violet Cell Staining (Optional)
- Micropipettes with capability of measuring volumes ranging from 1 μ l to 1 ml
- **37% paraformaldehyde stock concentration or 8% paraformaldehyde from other sources**
- Deionized or sterile water
- Squirt bottle, multichannel pipette and/or automated microplate washer
- Computer software capable of generating or displaying logarithmic functions
- Absorbent papers or vacuum aspirator
- Test tubes or microfuge tubes capable of storing ≥ 1 ml
- Orbital shaker, Incubator, and Stimulants
- **Poly-L-Lysine or Poly-D-Lysine for suspension or loosely attaching cells**

HEALTH AND SAFETY PRECAUTIONS

- Reagents provided in this kit may be harmful if ingested, inhaled or absorbed through the skin.
- Fixing Solution contains formaldehyde. Formaldehyde is known to be a highly toxic reagent. Personal protection is strongly recommended while working with this chemical.
- Stop Solution contains 2 N Sulfuric Acid (H_2SO_4) and is an extremely corrosive agent. Please wear proper eye, hand and face protection when handling this material. When the experiment is finished, be sure to rinse the plate with copious amounts of running water to dilute the Stop Solution prior to disposing the plate or strips.
- Crystal Violet is an intense stain reagent. Avoid contact stain and clothing.

EXPERIMENT DESIGN

- 1) **Cell Line:** The cell line must express the target protein. This protocol can be used directly for adherent cells. For suspension cells and loosely attached cells, two steps are required: 1) Coat the plates with 100 μ l of 10 μ g/ml Poly-L-Lysine (Sigma Cat# P4832, not included) to each well of the 96-well plate for 30 minutes at 37°C before proceeding to Step 1 of Assay Protocol (on page 13). Use 8% formaldehyde to fix the cells on Step 5 of Assay Protocol.
- 2) **Cell Number and Sensitivity:** The number of cells plated onto the 96-well plates depends on the expression level of Dysferlin protein in the cells, cell size, treatment conditions and incubation time. The cells used for testing should be around 75-90% confluent. Typically for HeLa cells, seed 30,000 cells per well overnight for treatment the following day. The Dysferlin Colorimetric Cell-Based ELISA Kit can detect Dysferlin expression in as little as 5,000 HeLa cells.
- 3) **Cell Treatment:** The cells can be treated with inhibitors, activators, stimulators (ie. chemicals, proteins/peptides) or a combination of the substances listed above. The cells can be treated with UV and serum starvation to meet the needs of the end-user.
- 4) **Positive and Negative Controls:** Mouse Anti-GAPDH Antibody (included) should be used to detect the internal positive controls for normalization of OD values of the target protein. The negative controls are HRP-Conjugated Anti-Rabbit IgG Antibody and HRP-Conjugated Anti-Mouse IgG Antibody alone in different wells (without the primary antibodies). Both positive and negative controls should be performed in the same plate with the Dysferlin target experiments.
- 5) **Accuracy and Precision:** Each condition should be performed in duplicate or in triplicate.

ASSAY PROTOCOL

Note: Please read the whole manual before performing the experiment.

1. Seed 100 μ l of 20,000 adherent cells into each well in a 96-well plate.
Note: Optimal seeding concentrations should be determined by investigator and experimental goals.

For suspension cells and loosely attached cells:

- a. Coat plates evenly with 20 μ l of Poly-L-Lysine (Sigma Cat# P4832)
 - b. After 5 minutes, remove solution by aspiration and rinse once with tissue culture grade water.
 - c. Allow 2 hours to dry prior to seeding cells.
2. Incubate cells overnight or at least 6 hours at 37°C, 5% CO₂.
 3. Treat the cells as desired.
 4. Remove the cell culture medium *gently* and fix cells with 100 μ l of 8% Paraformaldehyde per well. Incubate for 20 minutes at room temperature (RT).

Note: 8% Paraformaldehyde solution is volatile. Wear appropriate personal protection equipment (mask, gloves and glasses) when using this chemical.

5. Aspirate the solution and add 300 μ l of 1x Wash Buffer into each well being used and gently shake for 2-3 mins on an orbital shaker. Repeat this process 3 times. After the last wash ensure no liquid remains by inverting the plate and tapping it against clean paper towels.

Note: The plate can be stored at 4°C for a week after this step.

6. Add 100 μ l Quenching Buffer to each well and incubate for 20 mins at RT.
7. Repeat step 5.
8. Add 200 μ l of Blocking Buffer and incubate for 1 hour at RT.
9. Add 50 μ l of 1x Primary Antibody solutions into each well and incubate overnight at 4°C. If target concentration is known to be high, incubate for 2 hours at RT. Designate 2 wells for negative control by omitting the 1x Primary Antibodies.
10. Repeat step 5.
11. Add 50 μ l of HRP-Conjugated Anti-Rabbit IgG Antibody into the wells incubated with 1x Anti-Dysferlin Antibody.
12. Add 50 μ l of HRP-Conjugated Anti-Mouse IgG Antibody into the wells incubated with 1x Anti-GAPDH Antibody.

13. Incubate the plate for 1.5 hours at RT.
 14. Repeat step 5.
 15. Add 50 μ l of Ready to use Substrate into each well, cover plate from light, and incubate for 15-20 mins at RT.
 16. Add 100 μ l of Stop Solution into each well and read at 450 nm.
 17. Repeat step 5 and allow plate to air dry for 5 mins at RT.
 18. Add 50 μ l of Crystal Violet Solution into each well and incubate for 30 mins at RT.
- Note:** Crystal Violet is an intense stain. Avoid contact with skin and clothing.
19. Dip the plate into a bucket of water in the sink while keeping the water running, and carefully rinse the wells with ddH₂O until no more color is visible. Let the plate to dry for 30 mins.
 20. Add 100 μ l of SDS Solution into each well and incubate for 1 hour at RT.
 21. Read absorbance at 595 nm with microplate reader. If absorbance is too high, the solubilized Crystal Violet Solution may be diluted tenfold with ddH₂O on a separate 96-well plate.

DATA ANALYSIS

Background subtraction

Average the duplicate or triplicate absorbance readings for each control and sample, subtracting them from the averaged absorbance for wells where primary antibodies have been omitted.

Anti-GAPDH Internal positive control

The OD450 values obtained ensure the assay is functioning, and cell densities between wells should be proportionate to seeding concentration. Stimulation of cells should impact target primary antibody absorbance values, with no effect on the anti-GAPDH absorbance.

Crystal Violet Staining Normalization

The Crystal Violet staining method enables intensity normalization within the same well. By using the ratio, the OD450 readings obtained can be normalized with the OD595 values. This constitutes a within-well method of analysis.

If the absorbance was too high and a tenfold dilution was required. Multiply all values by 10 prior to normalization.

OD450 anti-Dysferlin/OD595 Crystal Violet.

By doing so, the expression levels of Dysferlin are adjusted to account for the cell density present in each well.

Stimulated to Non-Stimulated Comparison

After normalization Crystal Violet staining, the resultant proportional values can be used to analyze the effects of stimulants in this assay.

SHORT PROTOCOL

Seed cells into wells and incubate overnight at 37°C, 5% CO₂



Apply desired treatment conditions



Add 100ul of Fixing Solution and incubate 20 minutes at room temperature



Add 100ul of Quenching Buffer and incubate 20 minutes at room temperature



Add 200ul of Blocking Buffer and incubate for 1 hour at room temperature



Add 50ul of 1x Primary Antibodies and incubate overnight at 4°C



Add 50ul of HRP-Conjugated Secondary Antibodies and incubate for 1.5 hours at room temperature



Add 50ul of Ready-to Use Substrate and incubate for 30 minutes at room temperature



Add 50ul of Stop Solution and read OD at 450nm



TROUBLESHOOTING GUIDE

Problem	Cause	Solution
Low Signal	Storage and Expiration	Read Manual for storage condition and expiration.
	Antibody Dilution	Dilute antibody 1:100 with Antibody Diluent.
	Low Cell Number	Seed more cells.
	Cells Detach from Well-Bottoms	Add solutions slowly from the side wall of the wells. Use Poly-L-Lysine to coat the plate period seeding the cells, if necessary.
High Background	Washing	Remove Wash Buffer completely.
	High Cell Number	Reduce the amount of seeded cells.
Variation	Pipetting	Check and/or calibrate pipettes.
	Washing	Remove Wash Buffer completely.
	Cells Detach from Well-Bottoms	Add solutions slowly from the side wall of the wells. Use Poly-L-Lysine to coat the plate period seeding the cells, if necessary.

Important Note: During shipment, small volumes of product will occasionally become entrapped in the seal of the product vial. We recommend briefly centrifuging the vial to dislodge any liquid in the container's cap prior to opening.

Warning: This reagent may contain sodium azide and sulfuric acid. The chemical, physical, and toxicological properties of these materials have not been thoroughly investigated. Standard Laboratory Practices should be followed. Avoid skin and eye contact, inhalation, and ingestion. Sodium azide forms hydrazoic acid under acidic conditions and may react with lead or copper plumbing to form highly explosive metal azides. On disposal, flush with large volumes of water to prevent accumulation.

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