LSBio
an Absolute Biotech Company

# Human CCL4L1 ELISA Kit

# (Sandwich ELISA)

# User Manual

Catalog No. LS-F26793

It is important that you read this entire manual carefully before starting your experiment.

SAMPLE ONLY

This kit is for **Research Use Only**.
This kit is not approved for use in dogs or for clinical diagnosis.

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# ASSAY SPECIFICATIONS

**Target**: CCL4L1

**Synonyms**: CCL4L1, chemokine (C-C motif) ligand 4-like 1, AT744.2, CCL4L, LAG1, LAG-1, SCYA4L, SCYA4L2, MIP-1-beta, SCYA4L1, C-C motif chemokine 4-like, lymphocyte activation gene 1, macrophage inflammatory protein-1b2, small inducible cytokine A4-like

**Specificity**: This kit is for the detection of Human CCL4L1. No significant cross-reactivity or interference between CCL4L1 and analogs was observed. This claim is limited by existing techniques therefore cross-reactivity may exist with untested analogs.

**Sample Types**: This kit is recommended for use with Human Plasma and Serum. Use with other sample types is not supported.

**Detection:** Colorimetric - 450nm (TMB)

**Measurement:** Quantitative

**Detection Range**: 100–2500 pg/ml

**Sensitivity**: Typically less than 1 pg/ml

**Performance**: Intra-Assay CV (<9%); Inter-Assay CV (<10%)

**Limitations**: This kit is for **Research Use Only** and is not intended for diagnostic use. This kit is not approved for use in humans or for clinical diagnosis.

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## ASSAY PRINCIPLE

This assay is based on the sandwich ELISA principle. Each well of the supplied microtiter plate has been pre-coated with a target specific capture antibody. Standards or samples are added to the wells and the target antigen binds to the capture antibody. A Horseradish Peroxidase (HRP)-conjugated detection antibody is then added which binds to the captured antigen. Unbound antigen and detection antibody is washed away. A TMB substrate is then added which reacts with the HRP enzyme resulting in color development. A sulfuric acid stop solution is added to terminate color development reaction and then the optical density (OD) of the well is measured at a wavelength of 450 nm ± 2 nm. An OD standard curve is generated using known antigen concentrations; the OD of an unknown sample can then be compared to the standard curve in order to determine its antigen concentration.

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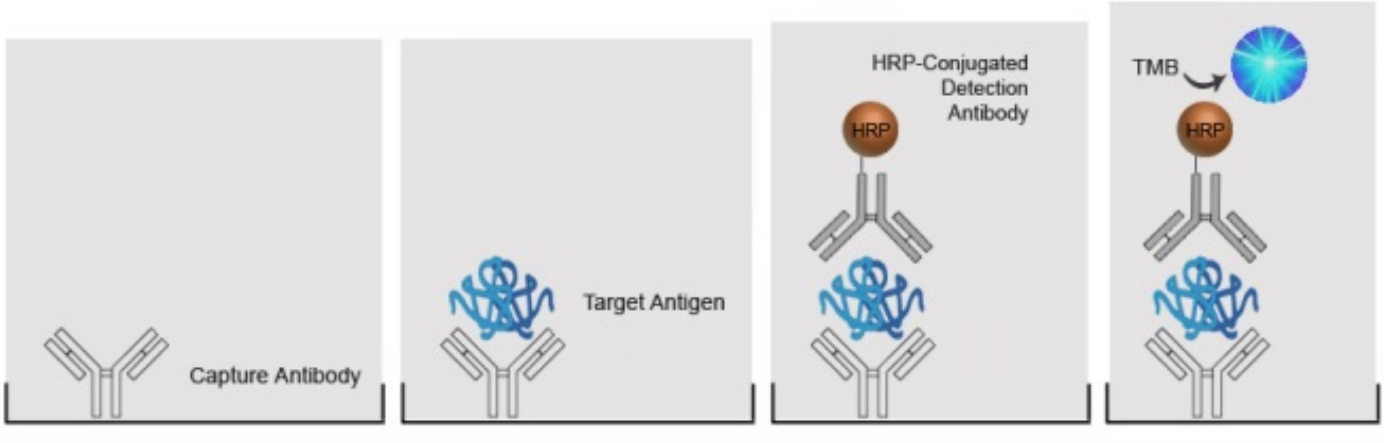
## Assay Principle Image

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## Kit Components and Storage

| Component | Quantity |
|---|---|
| Coated 96-well Strip Plate | 1 |
| Standard A (0pg/ml) | 1 vial |
| Standard B (100pg/ml) | 1 vial |
| Standard C (250pg/ml) | 1 vial |
| Standard D (500pg/ml) | 1 vial |
| Standard E (1000pg/ml) | 1 vial |
| Standard F (2500pg/ml) | 1 vial |
| Balance Solution | 1 vial x 3mL |
| HRP-Conjugate | 1 vial x 10mL |
| Wash Buffer (100x) | 1 vial x 10 mL |
| Substrate A | 1 vial x 6mL |
| Substrate B | 1 vial x 6mL |
| Stop Solution | 1 vial x 6mL |
| Adhesive Plate Sealers | 4 |
| Instruction Manual | 1 |

## Kit Storage

The unopened kit can be stored at 2°C to 8°C through the expiration date. Once opened, the kit can be stored at 2°C to 8°C for 1 month. Unused strips should be kept in a sealed bag with the desiccant provided to minimize exposure to damp air.

## Other Required Supplies

- Microplate reader with 450nm wavelength filter.
- An incubator which can provide stable incubation conditions up to 37°C±0.5°C.
- High-precision pipette and sterile pipette tips
- Eppendorf tubes
- Deionized or distilled water
- Absorbent paper

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# EXPERIMENTAL LAYOUT

The following is an example of how to layout a study. A dilution series of the positive control Standard should be run in duplicate or triplicate, with the last well in each series being the negative control blank. Samples should also be run in duplicate or triplicate. Unknown samples should be run as a dilution series in order to identify the optimal dilution that produces an OD reading within the OD range of the positive control Standard dilution series.

**Example 1:** Standard Curve and dilution series of an unknown sample.

| | 1 | 2 | 3 | 4 | ... |
|---|---|---|---|---|---|
| A | Standard A | Standard A | Sample (1:1) | Sample (1:1) | ... |
| B | Standard B | Standard B | Sample (1:10) | Sample (1:10) | ... |
| C | Standard C | Standard C | Sample (1:100) | Sample (1:100) | ... |
| D | Standard D | Standard D | Sample (1:1k) | Sample (1:1k) | ... |
| E | Standard E | Standard E | Sample (1:10k) | Sample (1:10k) | ... |
| F | Standard F | Standard F | Sample (1:100k) | Sample (1:100k) | ... |
| G | Blank | Blank | Sample (1:1,000k) | Sample (1:1,000k) | ... |
| H | Blank | Blank | Sample (1:10,000k) | Sample (1:10,000k) | ... |

**Example 2:** Standard Curve and samples run in duplicate.

| | 1 | 2 | 3 | 4 | ... |
|---|---|---|---|---|---|
| A | Standard A | Standard A | Sample A | Sample E | ... |
| B | Standard B | Standard B | Sample A | Sample E | ... |
| C | Standard C | Standard C | Sample B | Sample F | ... |
| D | Standard D | Standard D | Sample B | Sample F | ... |
| E | Standard E | Standard E | Sample C | Sample G | ... |
| F | Standard F | Standard F | Sample C | Sample G | ... |
| G | Blank | Blank | Sample D | Sample H | ... |
| H | Blank | Blank | Sample D | Sample H | ... |

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## SAMPLE COLLECTION

This assay is recommended for use with Human Plasma and Serum. Use with other sample types is not supported. The sample collection protocols below have been provided for your reference.

**Breast Milk** - Centrifuge samples for 20 minutes at 1,000 × g to remove particulates. Collect the supernatant for assaying. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Cell Lysates** - Collect and pellet the cells by centrifugation and remove the supernatant. Wash the cells three times with PBS*then resuspend in PBS*. Lyse the cells by ultrasonication four times. Alternatively freeze the cells to -20°C and thaw to room temperature three times. Centrifuge at 1,500 × g for 10 minutes at 2°C to 8°C to remove cellular debris. Collect the supernatant for assaying. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Erythrocyte Lysates** - Centrifuge whole blood for 20 minutes at 1,000 × g to pellet the cells and remove the supernatant. Wash the cells three times with PBS*then resuspend in PBS*. Freeze the cells freeze to -20°C and thaw to room temperature three times. Centrifuge at 5,000 × g for 10 minutes at 2°C to 8°C to remove cellular debris. Collect the supernatant for assaying. Erythrocyte lysates must be diluted with Sample Dilute before running. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Plasma** - Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge samples for 15 minutes at 1,000 × g at 2°C to 8°C within 30 minutes of collection. Collect the supernatant for assaying. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Platelet-Poor Plasma** - Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge samples for 15 minutes at 1,000 × g at 2°C to 8°C within 30 minutes of collection. It is recommended to centrifuge samples for 10 minutes at 10,000 × g for complete platelet removal. Collect the supernatant for assaying. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Sperm and Seminal Plasma** – Allow semen to liquefy at room temperature or 37°C. After liquefaction, centrifuge at 2,000 × g for 10 to 15 minutes. Collect seminal plasma supernatant for assaying. Wash the precipitated protein three times with PBS* then resuspend in PBS*. Lyse

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the cells by ultrasonication then centrifuge at 2,000 × g for 10 to 15 minutes. Collect the supernatant for assaying. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Serum** - Use a serum separator tube and allow samples to clot for 2 hours at room temperature or overnight at 4°C before centrifugation for 20 minutes at approximately 1,000 × g. Collect the supernatant for assaying. Grossly hemolyzed samples are not suitable for use in this assay. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Tissue Homogenates** – Because preparation methods for tissue homogenates vary depending upon tissue type users should research tissue specific conditions independently. The following is one example only. Rinse tissues in PBS* to remove excess blood and weighed before homogenization. Finely minced tissues and homogenized them in 5mL to 10mL of PBS*with a glass homogenizer on ice. Lyse the cells by ultrasonication or freeze the cells to -20°C and thaw to room temperature three times. Centrifuge homogenate at 5,000 × g for 5 minutes. Collect the supernatant for assaying. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Urine** - Aseptically collect the first urine of the day, mid-stream, voided directly into a sterile container. Centrifuge to remove particulate matter. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

**Cell culture supernatants, cerebrospinal, follicular, and lung lavage fluids, saliva, sweat, tears, and other biological fluids -** Centrifuge samples for 20 minutes at 1,000 × g to remove particulates. Collect the supernatant for assaying. Store un-diluted samples at -20°C or below. Avoid repeated freeze-thaw cycles.

* 1xPBS (0.02 mol/L pH 7.0-7.2)

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## SAMPLE COLLECTION NOTES

1. LSBio recommends that samples are used immediately upon preparation.
2. Avoid repeated freeze/thaw cycles for all samples.
3. In the event that a sample type not listed above is intended to be used with the kit it is recommended that the customer conduct validation experiments in order to be confident in the results.
4. Due to chemical interference, the use of tissue or cell extraction samples prepared by chemical lysis buffers may result in inaccurate results.
5. Influenced by the factors including cell viability, cell number or sampling time, samples from cell culture supernatant may not be detected by the kit.
6. Samples should be brought to room temperature (18°C to 25°C) before performing the assay without the use of extra heating.
7. Sample concentrations should be predicted before being used in the assaying. If the sample concentration is not within the range of the standard curve users must determine the optimal sample dilutions for their particular experiments.
8. LSBio is responsible for the quality and performance of the kit components but is NOT responsible for the performance of customer supplied samples use with the kit.

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## Reagent Preparation

Bring all reagents to room temperature (18°C to 25°C) before use.

**1x Wash Buffer**: If crystals have formed in the concentrate warm to room temperature and mix it gently until crystals have completely dissolved. Prepare 1,000 mL of Working Wash Buffer by diluting the supplied 10 mL of 100x Wash Buffer Concentrate with 990 mL of deionized or distilled water. Wash Buffer can be stored at 4°C once prepared.

All other reagents are ready-to-use.

## Reagent Preparation Notes

1. All solutions prepared from concentrates are intended for one-time use. Do not reuse solutions.
2. Reagents may adhere to the tube wall or cap during transport; centrifuge tubes briefly before opening.
3. All solutions should be gently mixed prior to use.
4. To minimize imprecision caused by pipetting, ensure that pipettes are calibrated. Pipetting volumes of less than 10 µL is not recommended.
5. Substrate Solution is easily contaminated so sterility precautions should be taken. Substrate Solution should also be protected it from light.
6. Do not substitute reagents from one kit lot to another. Use only those reagents supplied within this kit.
7. Due to the antigen specificity of the antibodies used in this assay, native or recombinant proteins from other manufacturers may not be detected by this kit.

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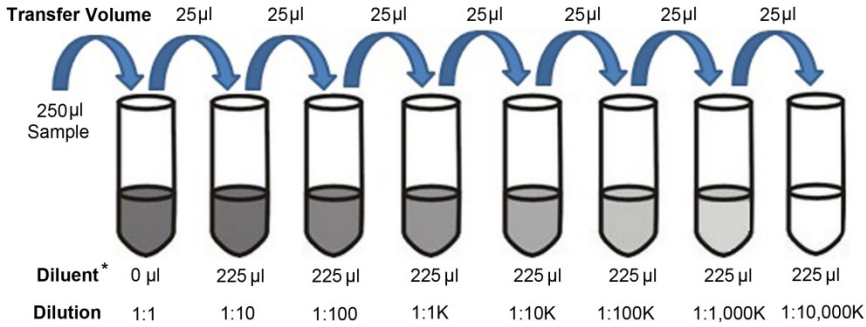
## SAMPLE PREPARATION

The resulting Optical Density (OD) values of your sample must fall within the OD values of the standard curve in order for the calculated antigen concentration to be accurate. In many cases samples will need to be diluted in order to lower the antigen concentration to sufficient levels. Information about antigen concentrations within various sample types may be available from the published literature; however it is often necessary to run a dilution series of each sample type. The following will prepare sufficient volumes to run the Sample dilution series in triplicate. In the case of small volume samples a preliminary step dilution, such as 1:5 or 1:10, can be made using PBS (0.02mol/L pH7.0-7.2) as the diluent. * **The final dilution should always be made using the same buffer that is used to dilute the Standards, and/or generate the Standard Curve.** Running duplicate or triplicate wells for each sample is recommended.




## STANDARDS PREPARATION

The Standards supplied with this kit are ready-to-use and no preparation is necessary.

## ASSAY PROCEDURE

Bring all reagents and samples to room temperature without additional heating and mixed thoroughly by gently swirling before pipetting (avoid foaming). Prepare all reagents and samples as directed in the previous sections.

1. Add 50 µL of PBS (0.02 mol/L pH 7.0-7.2) to a Blank well.
2. Add 5 µL of **Balance Solution** to each 50 µL sample of cell culture supernatant or tissue homogenate, if applicable, and mix well. Do NOT add to plasma or serum samples, standards, or the Blank.
3. Add 50 µL of **Standard** or **Sample** per well.
4. Add 100 µL of **HRP-conjugate** to each well (excluding the Blank well), cover with a plate sealer, and incubate for 1 hour at 37°C.
5. Aspirate the liquid from each well and wash five times. Wash by adding approximately 350 µL of **1x Wash Buffer** using a squirt bottle, multi-channel pipette, manifold dispenser, or automated washer. Allow each wash to sit for 10 seconds before completely aspirating. After the last wash, aspirate to remove any remaining Wash Buffer then invert the plate and tap against clean absorbent paper.
6. Add 50 µL of **Substrate A** and 50 µL of **Substrate B** to each well (including the Blank), gently agitate to ensure thorough mixing, and incubate **in the dark** for 15-20 minutes at 37 °C.
7. Add 50 µL of **Stop Solution** to each well. The blue color will change to yellow immediately. If color change does not appear uniform gently tap the plate to ensure thorough mixing. The Stop Solution should be added to wells in the same order and timing as was the substrate solution.
8. Determine the optical density (OD value) of each well immediately using a microplate reader set to 450 nm.

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## ASSAY PROCEDURE NOTES

1. **ELISA Plate**: Keep appropriate numbers of strips for one experiment and remove extra strips from microtiter plate. Removed strips should be placed in a sealed bag containing desiccant and stored at 4°C.
2. **Solutions**: To avoid cross-contamination change pipette tips between additions of each standard level, between sample additions, and between reagent additions. Also, use separate reservoirs for each reagent.
3. **Applying Solutions:** All solutions should be added to the bottom of the ELISA plate well. Avoid touching the inside wall of the well. Avoid foaming when possible.
4. **Assay Timing**: The interval between adding sample to the first and last wells should be minimized. Delays will increase the incubation time differential between wells which will significantly affect the experiment's accuracy and repeatability. For each step in the procedure total dispensing time for addition of reagents or samples should not exceed 10 minutes.
5. **Incubation**: To prevent evaporation and ensure accurate results, proper adhesion of plate sealers during incubation steps is necessary. Do not allow wells to sit uncovered for extended periods of time between incubation steps. Do not let wells dry out at any time during the assay. Strictly observe the recommended incubation times and temperatures.
6. **Washing**: Proper washing procedure is critical. Insufficient washing will result in poor precision and falsely elevated absorbance readings. Residual liquid in the reaction wells should be patted dry against absorbent paper during the washing process. Do not put absorbent paper directly into the reaction wells.
7. **Controlling Substrate Reaction Time**: After the addition of the Substrates periodically monitor the color development. Stop color development before the color become too deep by adding Stop Solution. Excessively strong color will result in inaccurate absorbance reading.
8. **Reading**: The microplate reader should be preheated and programmed prior to use. Prior to taking OD readings remove any

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residual liquid or fingerprints from the underside of the plate and confirm that there are no bubbles in the wells.

9. **Reaction Time Control**: Control reaction time should be strictly followed as outline.
10. **Stop Solution**: The Stop Solution is contains an acid, therefore proper precautions should be taken during its use, such as protection of the eyes, hands, face and clothing.
11. **Mixing**: During incubation times the use of a micro-oscillator at low frequency is recommended. Sufficient and gentle mixing is particularly important in producing reliable result.
12. Kits from different batches may differ slightly in detection range, sensitivity, and color developing time. Please perform the experiment exactly according to the supplied instructions.
13. Due to inter- and intra-assay variability it is recommended that appropriate carry over controls be included between assays.
14. Prior to running valuable samples LSBio recommends that the user runs a preliminary experiment using the supplied controls in order to validate the assay.
15. To minimize extra influence on the performance operation procedures and lab conditions, especially room temperature, air humidity, and incubator temperature, should be strictly controlled. It is also strongly suggested that the whole assay is performed by the same operator from the beginning to the end.
16. The kit should not be used beyond the expiration date on the kit label.

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# ASSAY PROCEDURE SUMMARY

Prepare all reagents, samples and standards.

Add 50 µL of PBS as a **Blank** well.

Add 5 µL of **Balance Solution** to each 50 µL sample of cell culture supernatant or tissue homogenate sample if applicable.

Add 50 µL of **Standard** or **Sample** to each well.

Add 100 µL of **1x HRP-conjugate** to each well (excluding the blank) and incubate for 1 hour at 37°C.

Aspirate and wash five times.

Add 50 µL of **Substrate A** and 50 µL **Substrate B** to each well and incubate for 15-20 minutes at 37°C.

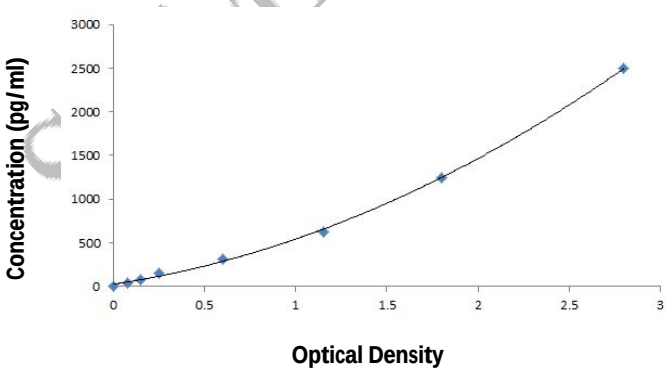
Add 50 µL of **Stop Solution**.

Read immediately at 450 nm.

## CALCULATION OF RESULTS

Average the duplicate readings for each standard, control, and sample and subtract the average zero standard optical density. Create a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the x-axis against the concentration on the y-axis and draw a best fit curve through the points on the graph. The data may be linearized by plotting the log of the target antigen concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. It is recommended to use some related commercial software to do this calculation, such as CurveExpert. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted the concentration read from the standard curve must be multiplied by the dilution factor.

**Typical Data:** The following standard curve is an example only and should not be used to calculate results for tested samples. A new standard curve must be generated for each set of samples tested.

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## TROUBLESHOOTING GUIDE

| Problem | Possible Cause | Solution |
|---|---|---|
| Poor standard curve | Inaccurate pipetting. | Check pipettes. |
| | Improper standard dilution. | Ensure the vial of standard has been briefly spun down and thoroughly dissolved by gentle mixing. |
| | Wells not completely aspirated. | Completely aspirate wells between steps. |
| Low signal | Too brief incubation times. | Ensure sufficient incubation time. |
| | Incorrect assay temperature. | Use recommended incubation temperature. Bring substrate to room temperature before use. |
| | Inadequate reagent volumes. | Check pipettes and ensure correct preparation. |
| | Improper dilution. | |
| Deep color but low value | Plate reader settings not optimal. | Verify the wavelength and filter setting in the plate reader. |
| | | Open the Plate Reader ahead to pre-heat. |

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## TROUBLESHOOTING GUIDE (CONTINUED)

| Problem | Possible Cause | Solution |
|---|---|---|
| Large CV | Inaccurate pipetting. | Check pipettes. |
| High background | Concentration of detector too high. | Use recommended dilution factor. |
| | Plate is insufficiently washed. | Review the manual for proper washing instructions. If using a plate washer, check that all ports are unobstructed. |
| | Contaminated wash buffer. | Make fresh wash buffer. |
| Low sensitivity | Improper storage of the ELISA kit. | All the reagents should be stored according to the instructions. |
| | Stop solution not added. | Stop solution should be added to each well before measurement. |

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**Important Note:** During shipment small volumes of product will occasionally become entrapped in the seal of the product vial. We recommend briefly centrifuging the vial to dislodge any liquid in the container's cap prior to opening.

**Warning**: This reagent may contain sodium azide and sulfuric acid. The chemical, physical, and toxicological properties of these materials have not been thoroughly investigated. Standard Laboratory Practices should be followed. Avoid skin and eye contact, inhalation, and ingestion. Sodium azide forms hydrazoic acid under acidic conditions and may react with lead or copper plumbing to form highly explosive metal azides. On disposal, flush with large volumes of water to prevent accumulation.

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