

Creatine Assay Kit (Colorimetric/Fluorometric)

LS-K300-100 (100 Tests) • Store at -20°C



Introduction

CREATINE is present in vertebrates and helps to supply energy to muscle. In humans and animals, approximately half of Creatine originates from food (mainly from fresh meat). Creatine supplementation has been investigated as a possible therapeutic approach for the treatment of muscular, neuromuscular, neurological and neurodegenerative diseases.

Simple, direct and automation-ready procedures for measuring Creatine are popular in research and drug discovery. LSBio's Creatine assay is based on enzymatic reactions leading to formation of a pink colored product. The optical density at 570 nm or fluorescence intensity at $\lambda_{em/ex}=590/530$ nm is directly proportional to the Creatine concentration in the sample.

Key Features

- High sensitivity and wide linear range. Use 10 μ L sample. Linear detection range 4 to 1000 μ M (colorimetric) or 0.5 to 50 μ M (fluorometric).
- Homogeneous and simple procedure. Simple "mix-and-measure" procedure allows reliable quantitation of Creatine within 30 minutes.

Applications

- Direct Assays: Creatine in biological samples (e.g. serum, plasma, urine, saliva etc).

Components

| Component | K300-100 |
|---------------------------|-------------|
| | 100 Tests |
| Assay Buffer | 20 mL |
| Enzyme A | 120 μ L |
| Enzyme B | 220 μ L |
| Dye Reagent | 220 μ L |
| Standard (20 mM Creatine) | 400 μ L |

Materials Not Supplied

Pipetting devices, and clear flat-bottom 96-well plates and optical density plate reader for colorimetric assays; black flat-bottom 96-well plate and fluorescence intensity plate reader for fluorometric assays.

Storage

The kit is shipped on ice. Store all kit components at -20 °C.

Assay Procedure

Sample preparation: SH-group containing reagents (e.g. mercaptoethanol, DTT) and EDTA may interfere with this assay and should be avoided in sample preparation. Solid samples can be extracted by homogenization in distilled water (dH₂O) and filtered or centrifuged. Liquid samples (e.g. serum, plasma and urine) can be assayed directly.

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Colorimetric Procedure

- Standards and Samples. Equilibrate all components to room temperature. Briefly centrifuge tubes before opening. Prepare a 1000 μM Creatine Standard Premix by mixing 15 μL of the 20 mM Standard and 285 μL dH₂O. Dilute Standard as follows.

| No | Premix + dH ₂ O | Vol (μL) | Creatine (μM) |
|----|-------------------------------------|-----------------------|----------------------------|
| 1 | 100 μL + 0 μL | 100 | 1000 |
| 2 | 60 μL + 40 μL | 100 | 600 |
| 3 | 30 μL + 70 μL | 100 | 300 |
| 4 | 0 μL + 100 μL | 100 | 0 |

Transfer 10 μL standards into separate wells of a clear, flat-bottom 96- well plate.

Transfer 10 μL of each sample into two separate wells, one serving as a sample blank well (R_{BLANK}) and one as a sample well (R_{SAMPLE}).

- Enzyme Reaction. For each standard and sample well, prepare Working Reagent by mixing 90 μL Assay Buffer, 1 μL Enzyme A, 1 μL Enzyme B and 1 μL Dye Reagent. Add 90 μL Working Reagent to the four Standards and the Sample Wells. Prepare blank control reagent by mixing 90 μL Assay Buffer, 1 μL Enzyme B and 1 μL Dye Reagent (i.e. no Enzyme A). Add 90 μL Blank control reagent only to the Sample Blank Wells.

Tap plate to mix. Incubate 30 min at room temperature.

- Read OD_{570nm}

Fluorometric Procedure

The fluorometric procedure is the same as for the colorimetric assay, except that (1) the detection range is up to 50 μM Creatine and (2) a black, flat-bottom 96-well plate is used. Creatine standards of 0, 15, 30 and 50 μM are prepared.

After incubation for 30 min at room temperature, read fluorescence intensity at $\lambda_{\text{ex}} = 530 \text{ nm}$ and $\lambda_{\text{em}} = 590 \text{ nm}$.

Calculations

Subtract the standard values from the blank value (#4) and plot the ΔOD or ΔF against standard concentrations. Determine the slope and calculate the Creatine concentration of Sample,

R_{SAMPLE} and R_{BLANK} are optical density or fluorescence intensity readings of the Sample and Sample Blank, respectively. n is the sample dilution fact.

$$[\text{Creatine}] = \frac{R_{\text{SAMPLE}} - R_{\text{BLANK}}}{\text{Slope } (\mu\text{M}^{-1})} \times n \quad (\mu\text{M})$$

Note: if the calculated Creatine concentration is higher than 1000 μM in the colorimetric assay or 50 μM in the fluorometric assay, dilute sample in dH₂O and repeat assay. Multiply result by the dilution factor n .

Conversions: 1000 μM Creatine equals 13.1 mg/dL or 131 ppm.

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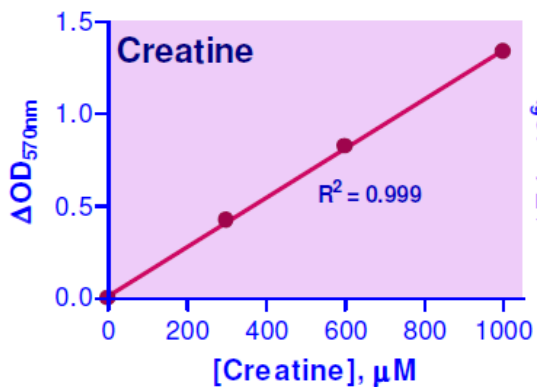
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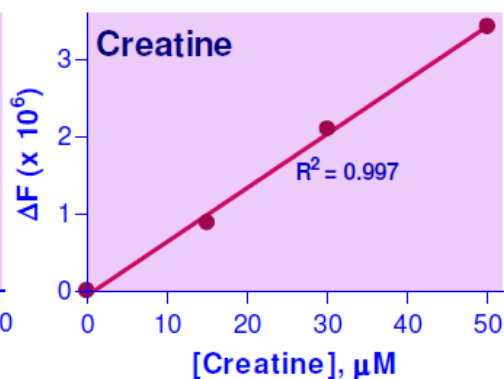
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Sample Data



96-well colorimetric assay



96-well fluorimetric assay

Version: V.08.09.2018

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