Xanthine Oxidase Assay Kit (Colorimetric/Fluorometric)

LS-K201-100 (100 Tests) • **Store at -20°C**

Introduction

Xanthine Oxidase catalyzes the oxidation of xanthine to uric acid. In addition, xanthine oxidase can catalyze the oxidation of hypoxanthine to xanthine, act on certain purines and aldehydes, and in certain cases produce the superoxide ion. Clinically, xanthine oxidase activity in blood can act as a marker for influenza, liver damage, and possibly cardiovascular health. Simple, direct and high-throughput assays for measuring xanthine oxidase activity find wide applications in research and drug discovery. This xanthine oxidase assay kit uses a single Working Reagent that combines the xanthine oxidase reaction and color reaction in one step. The change in color intensity of the reaction product at 570 nm or fluorescence intensity at $\lambda_{ex/em}$ = 530/585 nm is directly proportional to xanthine oxidase activity in the sample.

Key Features

- Sensitive and accurate. Use as little as 10 µL samples. Linear detection range in 96-well plate for 20 minute incubation: 0.03 to 25 U/L xanthine oxidase for colorimetric assays and 0.01 to 2.5 U/L for fluorometric assays.
- Simple and high-throughput. The procedure involves addition of a single working reagent and incubation for 20 min at room temperature.

Applications

- Direct Assays: xanthine oxidase activity in cell lysate, serum, and other biological samples.
- Drug Discovery/Pharmacology: effects of drugs on xanthine oxidase metabolism.

Components

Component	K201-100
	100 Tests
Assay Buffer	10 mL
HRP Enzyme	120 µL
Xanthine (5 mM Xanthine)	1.5 mL
Dye Reagent	120 µL
Standard (3% H_2O_2)	100 µL

Materials Not Supplied

Pipetting devices, centrifuge tubes, clear flat-bottom 96-well plates (e.g. VWR cat# 82050-760), black 96-well plates (e.g. Greiner Bio-One, cat# 655900) and plate reader.

Storage

The kit is shipped on ice. Store all reagents at -20°C. Shelf life: 6 months after receipt.

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Assay Procedure

Colorimetric Procedure

Samples can be analyzed immediately after collection, or stored in aliquots at –20°C. Avoid repeated freeze-thaw cycles. If particulates are present, centrifuge sample and use clear supernatant for assay.

1. Equilibrate all components to room temperature. During experiment, keep thawed Enzyme in a refrigerator or on ice.
2. H_2O_2 Standard Curve. Mix 5 µL 3% H_2O_2 and 914 µL dH_2O (final 4.8 mM) then mix 20 µL of the 4.8 mM H_2O_2 with 220 µL dH_2O to yield 400 µM H_2O_2. Prepare standards as shown in the Table below.

No	400 µM H_2O_2 + H_2O	Vol (µL)	H_2O_2 (µM)
1	100 µL + 0 µL	100	400
2	60 µL + 40 µL	100	240
3	30 µL + 70 µL	100	120
4	0 µL + 100 µL	100	0

   Transfer 10 µL standards and samples into separate wells.
3. Working Reagent. Prepare bulk working reagent by mixing 85 µL Assay Buffer, 10 µL 5 mM Xanthine, 1 µL HRP Enzyme (vortex briefly before pipetting), and 1 µL Dye Reagent per reaction well in a clean tube. Transfer 90 µL Working Reagent into each reaction well. Tap plate to mix.
4. Read optical density immediately (OD_0) at 570 nm (550-585 nm). Incubate 20 min at room temperature, and then read optical density again (OD_{20}).

Fluorometric Procedure

For fluorometric assays, the linear detection range is 0.01 to 2.5 U/L xanthine oxidase. Dilute the standards from Colorimetric Procedure 10x with dH_2O to obtain standards at 40, 24, 12 and 0 µM H_2O_2.

Transfer 10 µL standards and 10 µL samples into separate wells of a black 96-well plate.

Add 90 µL Working Reagent (see Colorimetric Procedure), tap plate to mix.

Read fluorescence immediately (F_0) at $\lambda_{ex/em}$ = 530/585 nm, incubate 20 min at room temperature, and then read fluorescence again (F_{20}).

Calculations

Subtract blank OD_{20} or F_{20} (water, #4) from all standard OD_{20} or F_{20} values and plot the ΔOD or ΔF against standard concentrations. Determine the slope using linear regression. Calculate the ΔOD_{Sample} or ΔF_{Sample} of all samples by subtracting OD_0 or F_0 from OD_{20} or F_{20} for each sample. Do the same for the blank (water, standard #4) to get ΔOD_{Blank} or ΔF_{Blank}. Calculate the activity using the equation below:

$$\text{XO Activity} = \frac{\Delta R_{SAMPLE} - \Delta R_{BLANK}}{\text{Slope } (\mu M^{-1}) \bullet t} \times n \quad \text{(U/L)}$$

Where ΔR_{Sample} and ΔR_{Blank} are the change in optical density or fluorescent values of the sample and blank, respectively. Slope is the slope of the H_2O_2 standard curve, *t* is the incubation time (20 minutes), and *n* is the dilution factor.

Notes: If the calculated sample XO activity is higher than 25 U/L in colorimetric assay or 2.5 U/L in fluorometric assay, dilute sample in water and repeat the assay. Multiply result by the dilution factor (*n*). For samples with low Xanthine Oxidase activity, the incubation time can be increased to up to 2 hours.

Unit definition: 1 U/L of Xanthine Oxidase catalyzes the conversion of 1 µmole of Xanthine to uric acid per minute at pH 7.0 and room temperature.

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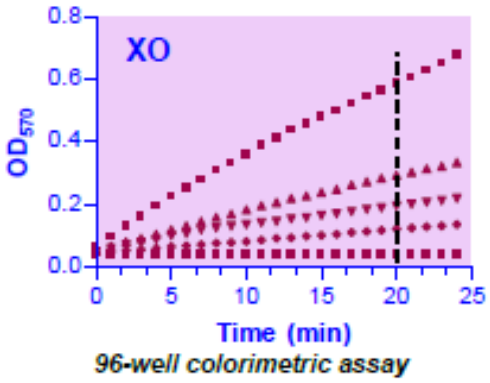
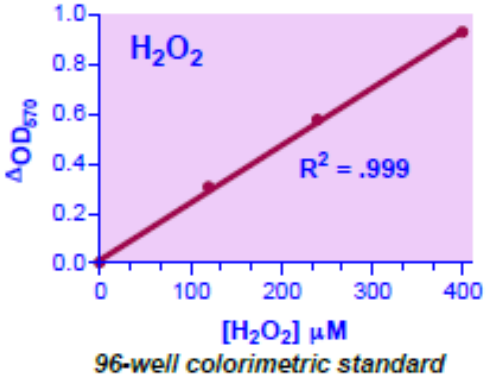
Sample Data



96-well colorimetric assay



96-well colorimetric standard

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