Acetate Assay Kit (Colorimetric/Fluorometric)

LS-K177-100 (100 Tests) • Store at -20°C



Introduction

ACETATE is a common anion and fundamental to all forms of life. When bound to coenzyme A, it is central to the metabolism of carbohydrates and fats. Its acid form, acetic acid, is produced and excreted by acetic acid bacteria, such as Acetobacter genus and Clostridium acetobutylicum, which are found universally in foodstuffs, water, and soil. Acetic acid is also a component of the vaginal lubrication of humans and other primates, where it appears to serve as a mild antibacterial agent. Acetic acid is the main component of vinegar, and extensively used in food, dyes, paints, glue and synthetic fibers.

LSBio's assay uses enzyme-coupled reactions to form a colored, fluorescent product. The color absorbance at 570nm or fluorescence intensity at 530nm/585nm is directly proportional to the acetate concentration in the sample.

Key Features

• Use as little as 10 μL samples. Detection range: 0.20 to 20 mM acetate for colorimetric assays and 0.13 to 2 mM for fluorometric assays.

Applications

- Direct Assays: acetate in biological samples such as serum/plasma, in food, agriculture and environmental samples.
- Drug Discovery/Pharmacology: effects of drugs on acetate metabolism.

Components

	K177-100	
Component	100 Tests	
Assay Buffer	25 mL	
Developer	1 mL	
Dye Reagent	120 μL	
АТР	120 μL	
Enzyme A	Dried	
Emzyme B	Dried	
Standard	1 mL	

Materials Not Supplied

Pipetting devices, centrifuge tubes, clear flat-bottom 96-well plates, optical density plate reader; black 96-well plates and fluorescence plate reader.

Storage

The kit is shipped on ice. Store all kit components at -20 °C.

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Assay Procedure

Fluorometric Procedure

Sample treatment: serum and plasma samples can be assayed directly. Acetic acid containing samples such as vinegars should be diluted in the Assay Buffer prior to assay. Samples should be clear, and free of precipitate or particles. If present, precipitate or particles should be removed by filtration or centrifugation.

- Equilibrate all components to room temperature. Briefly centrifuge tubes. Reconstitute Enzyme A with 600 μL Developer and Enzyme B with 120 μL Assay Buffer. Make sure both enzymes are fully dissolved before proceeding with the assay. During the experiment, keep Enzymes in a refrigerator or on ice. Reconstituted Enzyme A and Enzyme B are stable for four weeks if stored at -20 °C.
- 2. Standards and samples: prepare 400 μ L 2 mM Standard by mixing 4 μ L 200 mM standard with 396 μ L dH2O. Dilute standard in dH2O as follows.

No	$2 \text{ mM STD} + \text{H}_2\text{O}$	Vol (μL)	Acetate (mM)
1	100 μL + 0 μL	100	2.0
2	75 μL + 25 μL	100	1.5
3	50 μL + 50 μL	100	1.0
4	25 μL + 75 μL	100	0.5
5	0 μL +100 μL	100	0

Transfer 10 µL standards and 10 µL samples into separate wells of a black flat-bottom 96-well plate.

 Reaction. Prepare Working Reagent, for each reaction well, by mixing 90 μL Assay Buffer, 5 μL Enzyme A, 1 μL Enzyme B, 1 μL Dye Reagent and 1 μL ATP. Note: the Working Reagent should be prepared freshly and used within 20 min.

Transfer 90 µL Working Reagent to each well. Mix immediately and incubate for 30 min at room temperature. Read fluorescence intensity lex/lem = 530/585nm

Colorimetric Procedure

For colorimetric assays, the detection range is 0 to 20 mM acetate. Prepare 0, 4, 8, 12, 16 and 20 mM acetate standards in dH2O. Perform the assay the same as for Fluorometric Procedure, but use a clear flat bottom 96-well plate and read OD 570nm (550-585nm).

Notes: If the calculated acetate concentration of a sample is higher than 2 mM in fluorometric assay or 20 mM in colorimetric assay, dilute sample in water and repeat the assay. Multiply result by the dilution factor n.

Calculations

Subtract the water blank (Std #5) value from all the standard and sample values. Plot the DF or DOD of the standards against the standard concentrations. Determine the acetate concentration of samples from the standard curve.

Conversions: 1 mM acetate equals 5.9 mg/dL, 0.0059% or 59 ppm.

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Sample Data



Version: V.08.09.2018

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